

Original Investigation

Rare and Coding Region Genetic Variants Associated With Risk of Ischemic Stroke

The NHLBI Exome Sequence Project

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IMPORTANCE Stroke is the second leading cause of death and the third leading cause of years of life lost. Genetic factors contribute to stroke prevalence, and candidate gene and genome-wide association studies (GWAS) have identified variants associated with ischemic stroke risk. These variants often have small effects without obvious biological significance. Exome sequencing may discover predicted protein-altering variants with a potentially large effect on ischemic stroke risk.

OBJECTIVE To investigate the contribution of rare and common genetic variants to ischemic stroke risk by targeting the protein-coding regions of the human genome.


DESIGN, SETTING, AND PARTICIPANTS The National Heart, Lung, and Blood Institute (NHLBI) Exome Sequencing Project (ESP) analyzed approximately 6000 participants from numerous cohorts of European and African ancestry. For discovery, 365 cases of ischemic stroke (small-vessel and large-vessel subtypes) and 809 European ancestry controls were sequenced; for replication, 47 affected sibpairs concordant for stroke subtype and an African American case-control series were sequenced, with 1672 cases and 4509 European ancestry controls genotyped. The ESP's exome sequencing and genotyping started on January 1, 2010, and continued through June 30, 2012. Analyses were conducted on the full data set between July 12, 2012, and July 13, 2013.

MAIN OUTCOMES AND MEASURES Discovery of new variants or genes contributing to ischemic stroke risk and subtype (primary analysis) and determination of support for protein-coding variants contributing to risk in previously published candidate genes (secondary analysis).

RESULTS We identified 2 novel genes associated with an increased risk of ischemic stroke: a protein-coding variant in *PDE4DIP* (rs1778155; odds ratio, 2.15; $P = 2.63 \times 10^{-8}$) with an intracellular signal transduction mechanism and in *ACOT4* (rs35724886; odds ratio, 2.04; $P = 1.24 \times 10^{-7}$) with a fatty acid metabolism; confirmation of *PDE4DIP* was observed in affected sibpair families with large-vessel stroke subtype and in African Americans. Replication of protein-coding variants in candidate genes was observed for 2 previously reported GWAS associations: *ZFHX3* (cardioembolic stroke) and *ABCA1* (large-vessel stroke).

CONCLUSIONS AND RELEVANCE Exome sequencing discovered 2 novel genes and mechanisms, *PDE4DIP* and *ACOT4*, associated with increased risk for ischemic stroke. In addition, *ZFHX3* and *ABCA1* were discovered to have protein-coding variants associated with ischemic stroke. These results suggest that genetic variation in novel pathways contributes to ischemic stroke risk and serves as a target for prediction, prevention, and therapy.

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According to the 2010 Global Burden of Disease study, stroke is the second leading cause of death and the third leading cause of years of life lost.^{1,2} Ischemic stroke is the overt symptomatic manifestation of brain infarction, but the burden of cerebrovascular disease is much greater, with many more covert infarctions not resulting in the diagnosis of clinical stroke. More than 7% of asymptomatic adults in the general population have radiographic evidence of brain infarction,³ with substantially higher rates in the elderly population.⁴ Although many effective treatments for stroke exist, novel strategies for stroke prediction, prevention, and therapy need to be found.

Epidemiologic and family studies^{5,6} support an inherited component to stroke risk. Family-based linkage studies, including Cerebral Autosomal Dominant Arteriopathy With Subcortical Infarcts and Leukoencephalopathy and Cerebral Autosomal Recessive Arteriopathy With Subcortical Infarcts and Leukoencephalopathy,⁷ identified rare forms of stroke. Ischemic stroke studies include large candidate gene association meta-analyses,^{8,9} mendelian randomization,¹⁰ studies of affected sibpair families,¹¹ and genome-wide association studies (GWAS).¹²⁻¹⁴ The joint International Stroke Genetics Consortium/Wellcome Trust Case Control Consortium 2 (WTCCC2) effort¹⁴ identified *HDAC9* (7p21.1; GenBank [NM_058176](#)) and confirmed associations of ischemic stroke with variants in *PITX2* (GenBank [KJ891816](#)) and *ZFHX3* (GenBank [NM_006885](#)) (cardioembolic stroke) and in 9p21 (large-vessel/atherosclerotic stroke). The Australian Stroke Genetics Collaborative, the WTCCC2, and the International Stroke Genetics Consortium have identified and replicated the chromosome 6p21.1 large-artery susceptibility locus.¹³ Discovering the missing heritability for ischemic stroke could provide critical insights into the cause of the disease, novel pathways, and therapeutic targets.¹⁵

Rare, protein-coding variation could contribute a burden of risk for ischemic stroke, attributable to multiple variants in a gene rather than any individual variant.^{16,17} Exome sequencing in ischemic stroke has been performed only as a pilot study.¹⁸ Herein, we present results from what we believe to be the first large-scale study of protein-coding region variants in ischemic stroke from the National Heart, Lung, and Blood Institute (NHLBI) Exome Sequencing Project (ESP). We describe analyses used to identify novel stroke susceptibility

genes, determine genes and variants that are specific to stroke subtypes, and establish replication of previous GWAS and candidate gene associations.

Methods

Participating Studies and Design

The NHLBI ESP ascertained samples of European American and African American ancestry. Data were obtained on NHLBI cohorts (Atherosclerosis Risk in Communities, Coronary Artery Risk Development in Young Adults, Cardiovascular Health Study, Framingham Heart Study, Jackson Heart Study, the Multi-Ethnic Study of Atherosclerosis, and the Women's Health Initiative [WHI]) and 2 National Institute of Neurological Disorders and Stroke studies (Siblings With Ischemic Stroke Study [SWISS] and Ischemic Stroke Genetic Study [ISGS]). Characteristics of the participants analyzed are presented in **Table 1**.

Discovery analysis by exome sequencing focused on small-vessel and large-vessel subtypes in unrelated persons of European ancestry. Replication was conducted in small- and large-vessel subtypes in SWISS and ESP African American individuals, as well as ExomeChip data from European American participants in the WHI that included cardioembolic subtypes. Exome sequencing was performed on 365 unrelated cases (327 cases from NHLBI cohorts and 38 cases from ISGS: 114 with large-vessel subtype, 249 with small-vessel subtype, and 2 undetermined), 47 affected sibpairs from SWISS (94 individuals, used in replication: 28 pairs concordant for small-vessel subtype and 19 pairs concordant for large-vessel subtype), and African Americans (83 cases and 317 controls, used in replication). ExomeChip genotyping was performed on 1672 unrelated WHI ischemic stroke cases: 107 with large-vessel subtype, 224 with small-vessel subtype, 605 with cardioembolic subtype, and 736 of undetermined subtype. Cases classified as *undetermined* were included in the overall case-control analysis but not the subtype-specific analyses. Controls (n = 809) were identified as participants with exome sequence data who had not experienced a stroke or myocardial infarction at baseline or on subsequent follow-up and as individuals who were originally ascertained on the basis of low blood pressure or decreased low-density lipoprotein chole-

Table 1. Characteristics of Samples Used in the Discovery and Replication Analyses

Sample	Cases						Controls	
	Female, No./Total No. (%)	Age Mean (SD), y	Type of Stroke, No. (%)				Female, No./Total No. (%)	Age, Mean (SD), y
			Small Vessel	Large Vessel	Cardioembolic	Undetermined		
EA ESP	312/365 (85.5)	62.4 (11.6)	249 (68.2)	114 (31.2)	0	2 (0.5)	460/809 (56.9)	58.5 (11.7)
AA ESP	68/83 (81.9)	57.0 (9.2)	57 (68.7)	23 (27.7)	0	3 (3.6)	215/317 (67.8)	59.3 (8.3)
SWISS	36/94 (38.3)	67.4 (11.5)	56 (59.6)	38 (40.4)	0	0	NA	NA
WHI ExomeChip	1672/1672 (100)	68.7 (5.8)	224 (13.4)	107 (6.4)	605 (36.2)	736 (44.0)	4509/4509 (100)	66.9 (6.4)

Abbreviations: AA, African ancestry; EA, European ancestry; ESP, Exome Sequencing Project; NA, not available; SWISS, Siblings With Ischemic Stroke Study; WHI, Women's Health Initiative.

terol levels. Controls for ExomeChip analysis were 4509 WHI participants who self-reported having no history of stroke and myocardial infarction and no family history of stroke.

Ischemic Stroke Classification

Ischemic stroke was defined as a typical clinical syndrome with radiologic confirmation of brain infarction or the absence of an alternative diagnosis for the clinical syndrome. Stroke subtyping was based on the Trial of Org 10172 in Acute Stroke Treatment classification system.¹⁹ Brain imaging by computed tomographic scanning, magnetic resonance imaging, or autopsy confirmation was available to classify all cases. Relevant institutional review boards approved participating studies, and all participants provided written informed consent for participating in the original study as well as for genetic research. There was no financial compensation for use of existing data and samples from the contributing cohorts as part of this study.

Genetic Data

Exome sequencing was performed at the Broad Institute and the University of Washington (eAppendix 2 in the [Supplement](#)). ExomeChip genotyping was performed at the Broad Institute (eAppendix 3 in the [Supplement](#)). The ESP exome sequencing and ExomeChip genotyping started on January 1, 2010, and continued through June 30, 2012. Analyses were conducted on the full data set between July 12, 2012, and July 13, 2013. A total of 225 239 protein-coding variants were common to both ESP exome sequencing and ExomeChip genotyping, of which 119 963 were polymorphic in the full ESP sample (n = 7355). The self-reported ancestry and genetic ancestry distributions by principal components analysis are shown in eFigure 1 in the [Supplement](#).

Statistical Analysis

Single-variant tests and 2 gene-based tests were used to determine the association with ischemic stroke, conditional on the minor allele frequency (MAF) of individual variants (eAppendix 4 in the [Supplement](#)). Variants with MAF of 0.5% or more and at least 100 observations with a nonmissing genotype were analyzed using a single-variant test. For gene-based tests, only missense, splice, or nonsense variants were considered. Variants with MAF of 5% or less were analyzed with the sequence kernel association test (SKAT),²⁰ or variants with MAF of 1% or less analyzed with the combined multivariate and collapsing method, using variants with MAF of less than 1% (CMC/T1 test).²¹ For SKAT and CMC/T1, only genes with cumulative MAF of 0.5% or more were assessed. Analyses were conducted separately for exome sequence (combining data from all cohorts and ISGS) and ExomeChip (WHI) data; a meta-analysis was conducted using seqMeta software (<http://cran.r-project.org/web/packages/seqMeta/>).²² Covariate adjustment included participant's age at baseline and sex, as appropriate. For autosomal variants, a log-additive genetic model was used; for sex chromosome variants, a dominance model was used.

Ischemic stroke and subtype analyses used 2037 cases and 5318 controls with ESP exome sequencing or ExomeChip data.

Association of ischemic stroke with any single variant used $P < 4.17 \times 10^{-7}$ (0.05/119 963) as the threshold for exomewide significance. Restriction to variants with MAF of 0.005 or more and at least 100 observations with a nonmissing genotype yielded 25 467 variants, with a significance threshold at $P < 1.96 \times 10^{-6}$. Gene-level associations used a variety of statistical tests and MAF cutoffs²³; CMC/T1 is most powerful when all variants in a gene influence risk in the same direction; SKAT is more powerful when the variants in a gene influence risk in opposite directions. For CMC/T1, genes were included for individual variants with MAF of 0.01 or less and cumulative MAF of 0.005 or more, resulting in 6173 informative genes with a threshold of $P < 8.10 \times 10^{-6}$. For SKAT, there were 8361 genes with MAF of 0.05 or less and cumulative MAF of 0.005 or more, with a threshold of $P < 5.98 \times 10^{-6}$. In the discovery phase, these analyses were conducted for all ischemic stroke, small-vessel subtype, and large-vessel subtype. Affected sibpairs from SWISS were analyzed using identity-by-descent methods (eAppendix 4 in the [Supplement](#)). Regions of excess identity by descent were examined, focusing on missense, nonsense, stop-gain/loss functional variants. Single-variant and aggregate identity-by-descent probabilities of each gene across all available sibpairs were calculated to generate logarithm-of-the-odds scores for excess identity-by-descent sharing, adjusting for local recombination rates based on European ancestry estimates (HapMap phase 3), repeated for sibpairs concordant for small-vessel and large-vessel subtypes. The distribution of scores from the identity-by-descent analyses is shown in eFigure 2 in the [Supplement](#).

Targeted secondary analysis of 59 candidate genes previously associated with ischemic stroke, stroke risk factors, and mendelian syndromes yielded 134 variants. Most of these variants are not located in protein-coding regions; however, proxies were identified with strong linkage disequilibrium with ESP-defined protein-coding variants. For single-variant tests, significance was defined as $P < 1.86 \times 10^{-4}$. For gene-based tests, 33 genes contained polymorphic variants for analysis and significance was defined as $P < 7.58 \times 10^{-4}$ since 2 gene-based tests (CMC/T1 and SKAT) were used.

Results

Exome Variants Associated With Ischemic Stroke

Single-variant tests identified 2 protein-coding variants significantly associated with ischemic stroke ([Table 2](#)). A common (MAF, 0.316) missense variant, [rs1778155](#), in *PDE4DIP* (phosphodiesterase 4D-interacting protein; 1q21.1; GenBank [NM_014644](#)) was associated with ischemic stroke (odds ratio [OR], 2.15; $P = 2.63 \times 10^{-8}$). An infrequent (MAF, 0.017) missense variant, [rs35724886](#), in *ACOT4* (acyl-coenzyme A thioesterase 4; 14q24.3; GenBank [NM_152331](#)) was associated with ischemic stroke (OR, 2.04; $P = 1.2 \times 10^{-7}$). Although not reaching exomewide significance, a common (MAF, 0.379) variant, [rs5001076](#), in *PRIM2* (primase, DNA, polypeptide 2 [58 kilodaltons (kDa)]); 6p11.2; GenBank [NM_000947](#)) showed evidence of increased risk with ischemic stroke (OR, 2.11; $P = 2.96 \times 10^{-6}$) ([Table 2](#)).

Table 2. Association Results for Coding Region Variants and Risk of Ischemic Stroke With a Meta-analysis $P < 5 \times 10^{-6}$ Displayed

Gene	Location	Gene Size, kb	Associated SNV	MAF	OR			P Value			No. of SNVs in SKAT	SKAT Gene-Level P Value		
					Discovery	Repli-cation	Meta-analysis	Discovery	Repli-cation	Meta-analysis		Discovery	Repli-cation	Meta-analysis
<i>PDE4DIP</i>	1q21.1	240.030	rs1778155	0.316	2.147	NA	2.147	2.63×10^{-8}	NA	2.63×10^{-8}	42	.97	.24	.45
<i>ACOT4</i>	14q24.3	4.791	rs35724886	0.017	2.044	NA	2.044	1.24×10^{-7}	NA	1.24×10^{-7}	1	1.36×10^{-7}	NA	1.36×10^{-7}
<i>PRIM2</i>	6p11.2	333.773	rs5001076	0.379	2.105	NA	2.105	2.96×10^{-6}	NA	2.96×10^{-6}	4	7.2×10^{-5}	NA	7.2×10^{-5}

Abbreviations: kb, kilobase; MAF, minor allele frequency; OR, odds ratio; SNV, single-nucleotide variant; SKAT, sequence kernel association test.

Figure. Associations With Ischemic Stroke in *PDE4DIP*

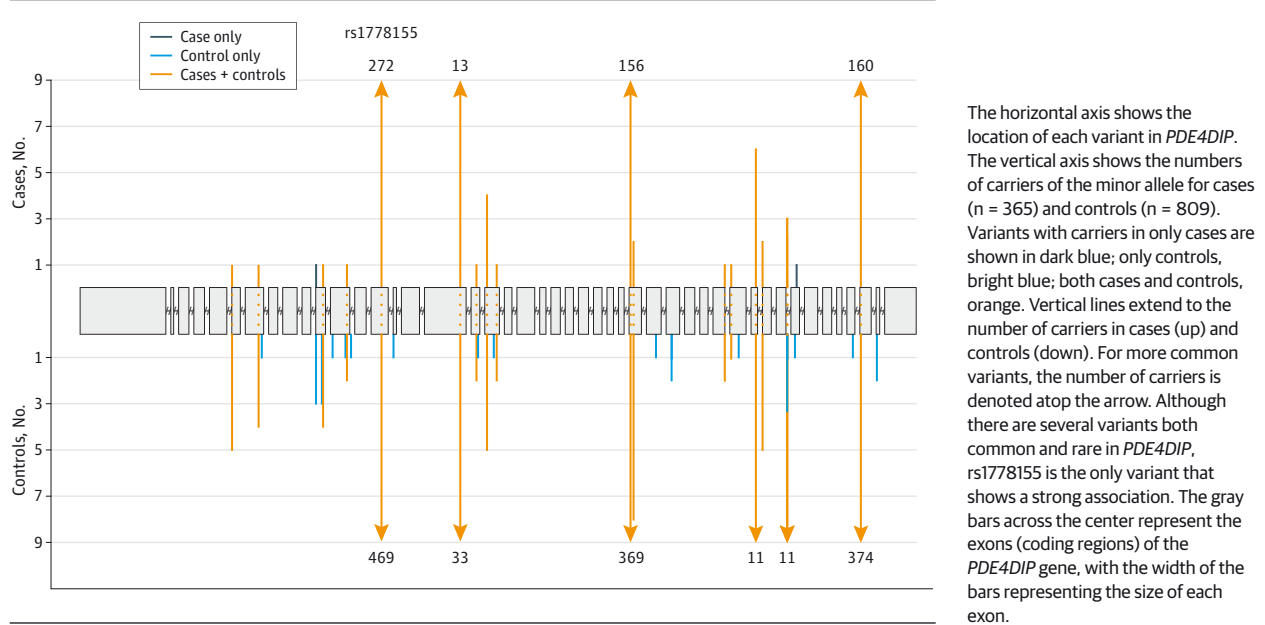


Table 3. Association Results for Coding Region Variants and Risk of Small-Vessel Ischemic Stroke With Meta-analysis $P < 5 \times 10^{-4}$ Displayed

Gene	Location	Gene Size, kb	SNV	MAF	Test Results			P Value		
					OR	P Value				
					ESP	Exome-Chip	Meta-analysis	ESP	Exome-Chip	Meta-analysis
Single-Variant Test Used										
<i>PDE4DIP</i>	1q21.1	240.030	Best, rs1778155	0.308	2.029	NA	2.029	7.96×10^{-6}	NA	7.96×10^{-6}
<i>ACOT4</i>	14q24.3	4.791	Best, rs35724886	0.018	1.952	NA	1.952	2.43×10^{-5}	NA	2.43×10^{-5}
Gene-Based Test Used (SKAT)										
<i>CEP164</i>	11q23.3	98.712	n = 24	Cumulative, 0.010	NA	NA	NA	.15	4.9×10^{-6}	7.93×10^{-6}

Abbreviations: kb, kilobase; MAF, minor allele frequency; NA, not applicable; OR, odds ratio; SKAT, sequence kernel association test; SNV, single-nucleotide variant.

Rare and infrequent protein-coding variants in *PDE4DIP* from exome sequencing in 365 cases and 809 controls and their estimated effect on ischemic stroke risk are shown in the Figure. A large complex gene (240.030 kilobases [kb]; 2346 amino acids, 265 kDa), *PDE4DIP* has 86 distinct introns, 64 different messenger RNAs, 49 alternatively spliced variants, and at least 10 transcription factor binding sites in its promoter. In contrast, *ACOT4* is a small gene (4.791 kb; 421 amino acids, 46 kDa) with 2 distinct introns

and 1 spliced messenger RNA. Both genes are expressed in the brain and in other tissues.

The results of no single-variant or gene-based test attained statistical significance for the small-vessel (lacunar) stroke subtype (Table 3). *PDE4DIP* rs1778155 had direction and size of effect on risk consistent in the all-stroke and small-vessel stroke categories (OR, 2.03; meta-analysis $P = 7.96 \times 10^{-6}$). *ACOT4* rs35724886 also had the same direction and similar size of effect on risk (OR, 1.95; meta-analysis

Table 4. Association Results for Coding Region Variants and Risk of Large-Vessel Ischemic Stroke With Meta-analysis $P < 5 \times 10^{-4}$ Displayed

Gene	Location	Gene Size, kb	SNV	MAF	Test Results			P Value		
					ESP-Sequence	Exome-Chip	Meta-analysis	ESP-Sequence	Exome-Chip	Meta-analysis
Single-Variant Test Used										
<i>SPSB3</i>	16p13.3	16.989	Best, rs147735377	0.006	NA	58.225	58.225	NA	9.62×10^{-6}	9.62×10^{-6}
<i>PDE4DIP</i>	1q21.1	240.030	Best, rs1778155	0.302	2.40	NA	2.403	2.18×10^{-5}	NA	2.18×10^{-5}
<i>DNAH14</i>	1q42.12	503.033	Best, rs41267347	0.080	2.33	2.03	2.181	.0016	.0058	3.06×10^{-5}
<i>MPDZ</i>	9p23	173.887	Best, rs188840960	0.009	2.51	30.35	8.497	.2056	4.45×10^{-6}	3.90×10^{-5}
<i>ZNF778</i>	16q24.3	11.855	Best, rs117690401	0.040	4.380	NA	4.380	4.88×10^{-5}	NA	4.88×10^{-5}
<i>SLC22A5</i>	5q31.1	25.906	Best, rs28383481	0.005	0.9289	172.89	16.853	.9439	4.25×10^{-8}	5.48×10^{-5}
<i>PGAP1</i>	2q33.1	94.793	Best, rs62185645	0.009	23.482	NA	23.482	6.01×10^{-5}	NA	6.01×10^{-5}
<i>KIF21A</i>	12q12	150.163	Best, rs78616703	0.005	2.44	84.71	16.290	0.3811	3.07×10^{-6}	6.06×10^{-5}
<i>MX1</i>	21q22.3	38.911	Best, rs140022520	0.005	51.953	NA	51.953	9.51×10^{-5}	NA	9.51×10^{-5}
<i>ACOT4</i>	14q24.3	4.791	Best, rs35724886	0.015	2.384	NA	2.384	1.07×10^{-4}	NA	1.07×10^{-4}
Gene-Based Test Used (SKAT)										
<i>SPSB3</i>	16p13.3	16.989	n = 3	Cumulative, 0.009	NA	NA	NA	.3169	1.05×10^{-5}	1.33×10^{-5}
<i>FAM151A</i>	1p32.3	14.380	n = 11	Cumulative, 0.053	NA	NA	NA	5.12×10^{-4}	.3479	4.20×10^{-5}
<i>MX1</i>	21q22.3	38.911	n = 8	Cumulative, 0.007	NA	NA	NA	5.20×10^{-5}	.0696	5.97×10^{-5}
<i>SLC22A5</i>	5q31.1	25.906	n = 11	Cumulative, 0.011	NA	NA	NA	.9600	6.05×10^{-8}	7.04×10^{-5}
<i>PGAP1</i>	2q33.1	94.793	n = 7	Cumulative, 0.013	NA	NA	NA	5.85×10^{-5}	.9556	8.96×10^{-5}
<i>KIF21A</i>	12q12	150.163	n = 15	Cumulative, 0.074	NA	NA	NA	.3302	1.05×10^{-6}	1.08×10^{-4}

Abbreviations: ESP, Exome Sequencing Project; kb, kilobase; MAF, minor allele frequency; NA, not applicable; OR, odds ratio; SNV, single-nucleotide variant; SKAT, sequence kernel association test.

$P = 2.43 \times 10^{-5}$). One novel gene, *CEP164* (centrosomal protein 164 kDa; 11q23.3; GenBank [NM_014956](#)) with 24 rare variants (cumulative MAF, 0.01) approached statistical significance (SKAT meta-analysis $P = 7.93 \times 10^{-6}$).

Within large-vessel (atherosclerotic) stroke, no single-variant or gene-based test attained statistical significance (Table 4). *PDE4DIP* rs1778155 had a strong effect and consistent direction (OR, 2.40; $P = 2.18 \times 10^{-5}$). Single-variant tests did not support *ACOT4* rs35724886, but gene-based tests of many infrequent or rare variants provided suggestive evidence for association (eg, *SPSB3* [GenBank [KJ899913](#)], *SLC22A5* [GenBank [KJ897580](#)], *PGAP1* [GenBank [NM_024989](#)], *KIF21A* [GenBank [NM_001173464](#)], and *MX1* [GenBank [NM_001144925](#)]) with large-vessel subtype.

Cases of cardioembolic ischemic stroke were not selected for ESP exome sequencing; these participants were sequenced only if they were selected for other (nonstroke) phenotypes or were included later for ExomeChip genotyping. No statistically significant associations with cardioembolic stroke were found by either single-variant or gene-based tests. The most significant single-variant association was with *PP1P5K2* rs35671301 (GenBank [NM_001276277](#)) (diphosphoinositol pen-

takisphosphate kinase 2; 5q21.1; OR, 3.59; MAF, 0.011; meta-analysis $P = 1.01 \times 10^{-5}$). The SKAT test also provided support for *PP1P5K2* on risk in the cardioembolic stroke (5 variants with cumulative MAF, 0.104; meta-analysis $P = 1.51 \times 10^{-5}$).

Replication of Exome Variants Associated With Ischemic Stroke

Single-nucleotide polymorphisms in *PDE4DIP* and *ACOT4* were analyzed in an independent set of affected sibpairs from SWISS and unrelated ESP African American participants. The *PDE4DIP* rs1778155 variant had significantly increased allele sharing identity by descent ($P = 1.16 \times 10^{-4}$) and was observed in 16 of 19 large-vessel concordant pairs with a logarithm of the odds of 6.07 (OR, 2.41; MAF, 0.302; $P = 2.18 \times 10^{-5}$) and a false discovery rate of $P = 6.39 \times 10^{-3}$. *ACOT4* rs35724886 was not polymorphic in either the SWISS or WHI ExomeChip data. A significant association with ischemic stroke was observed for *PDE4DIP* rs1778155 in the ESP African American samples (MAF, 0.191; OR, 2.34, $P = 7.50 \times 10^{-4}$). This variant failed genotype quality control metrics on the ExomeChip and was not analyzed. In African Americans, no significant association was detected for

ACOT4 rs35724886 with ischemic stroke although the effect size was similar (OR, 1.86; MAF, 0.180; $P = .31$).

Analysis of Previously Identified Candidate Genes and Variants

We identified 59 published candidate genes to be analyzed with the single-variant and gene-based tests. A single variant in *ABCA1* (adenosine triphosphate-binding cassette, subfamily A [*ABC1*], member 1; 9q31.1; GenBank [NM_005502](#)) was associated with all ischemic stroke (meta-analysis $P = 1.94 \times 10^{-4}$) and in both subtypes (small-vessel meta-analysis $P = .008$; large-vessel meta-analysis $P = .02$). Variants in *ZFHX3* (zinc finger homeobox 3; 16q22.3), a candidate gene for atrial fibrillation, were nominally associated with all ischemic stroke using the CMC/T1 test ($P = .001$), primarily in the cardioembolic subtype ($P = .04$). Results for all candidate genes are presented in eTables 1 through 4 in the [Supplement](#).

Discussion

We report the results of ESP exome sequencing and ExomeChip genotyping of 2037 cases of ischemic stroke and 5318 controls, with replication in an African American population and a collection of affected sibpairs. We present evidence supporting the role of protein-coding variants in 2 novel genes, *PDE4DIP* and *ACOT4*, which are associated with an increased risk of ischemic stroke. We evaluated protein-coding variants in previously reported genes associated with ischemic stroke, and support the role of *ABCA1* in large-vessel stroke and *ZFHX3* in cardioembolic stroke.

A primary interacting partner of *PDE4DIP* is *PDE4D* (phosphodiesterase 4D, cAMP-specific; GenBank [NM_001104631](#)), originally identified as linked to ischemic stroke in Icelandic families.²⁴ The *PDE4D* variants have been extensively studied²⁵⁻²⁸ for association with ischemic stroke risk in diverse populations, with equivocal results. A meta-analysis²⁹ of ischemic stroke was conducted on 7 studies, using approximately 12 000 cases and 15 000 controls that included 6 single-nucleotide polymorphisms in *PDE4D*. In the meta-analysis, *PDE4D* rs702553 had a significant association with ischemic stroke. The *PDE4D* result, coupled with *PDE4DIP* identified in the present study,^{30,31} supports a role of this pathway in ischemic stroke.

The *ACOT4* gene encodes an enzyme that catalyzes the hydrolysis of acyl-coenzyme A (CoA) to the free fatty acid and coenzyme A, regulating intracellular levels of acyl-CoA and fatty acids. Selected fatty acids are associated with adverse outcomes including dyslipidemia, inflammation, myocardial infarction, and other cardiovascular mortality.³²⁻³⁶ Human *ACOT4* hydrolyzes succinyl-CoA, glutaryl-CoA, and long-chain acyl-CoA reactions catalyzed by multiple enzymes in other species. Expression of *ACOT4* is regulated by peroxisome proliferator-activated receptor- α ,^{37,38} a nuclear receptor that has been implicated in vascular and cardiac disease and represents a therapeutic target in diabetes mellitus.³⁹ *ACOT4* links risk for ischemic stroke with fatty acid metabolism.

This study, which we believe to be the largest on exome sequencing in stroke, has limitations. The replication of *PDE4DIP* rs1778155 association with ischemic stroke succeeded in both affected sibpair families and a small case-control cohort of African ancestry; however, it was not replicated in ExomeChip analyses. Furthermore, the *ACOT4* rs35724886 infrequent variant was not informative in the replication samples and, therefore, was not replicated. Because these are protein-coding variants, their coverage in GWAS reports (eg, METASTROKE)⁴⁰ is often incomplete. *PDE4DIP* rs1778155 has no surrogate within r^2 value greater than 0.4 in the 1000 Genomes Project data; *ACOT4* rs35724886 has no surrogate within r^2 value greater than 0.8 in the *ACOT2/ACOT4/ACOT6* cluster. In the joint ImmunoChip/WTCCC2/METASTROKE data,⁴⁰ the *PDE4DIP* rs1778155 variant is 20.5 Mb proximal to the *SELP* (selectin P [granule membrane protein 140kDa, antigen CD62; 1q24.2]; GenBank [NM_003005](#)) rs3917792 variant (intronic, associated with all ischemic stroke: OR, 1.14; $P = 1.76 \times 10^{-7}$). However, *ACOT4* rs35724886 is only 1.4 Mb from the METASTROKE-identified *RGS6* (regulator of G-protein signaling 6; 14q24.2; GenBank [NM_001204416](#)) rs2238238 variant (intronic, associated with all ischemic stroke: OR, 1.18; $P = 1.65 \times 10^{-6}$). Thus, METASTROKE provides indirect support for the *ACOT4* contribution to ischemic stroke. Several candidate genes and loci were not analyzed, since few protein-coding variants were identified through ESP exome sequencing or included on the ExomeChip (eg, *HDAC9*, *PDE4D*). The 9p21.3 region, previously associated with large-vessel ischemic stroke, contains few protein-coding genes and, therefore, few coding-region variants. Only 3 variants in *CDKN2A* (cyclin-dependent kinase inhibitor 2A; GenBank [NG_007485](#)) and 1 variant in *CDKN2B-AS1* (CDKIN2B-antisense RNA 1; GenBank [HG975381](#)) were analyzed, with no variant associated with overall ischemic stroke or stroke subtype. Finally, our discovery sample focused on small- and large-vessel ischemic stroke subtypes to increase statistical power to detect novel genetic loci and reduce heterogeneity. Subsequent epidemiologically robust sampling approaches, consistent with the underlying distribution of subtypes in the population, should be conducted. Such studies would enable estimation of risk attributable to these genes (eg, *PDE4DIP* and *ACOT4*) and pathways to small- and large-vessel stroke and the effect of these variants in individual risk profiles for ischemic stroke.

The NHLBI ESP is the first large-scale exome sequencing project to focus on complex human phenotypes rather than mendelian disorders. Although data from both European Americans and African Americans were sequenced in the ESP, only European Americans had sufficiently large numbers of ischemic stroke to permit robust genetic analyses. The motivation for exploring the exome was that the protein-coding region variants would have greater likelihood of functional impact and, therefore, larger effects on disease risk, although examples of this remain few and require large sample sizes, similar to those of GWAS. Nevertheless, exome sequencing has, as exhibited in the present study, provided novel gene targets and biological pathways for examination on their role in ischemic stroke.

Conclusions

Our study has led to the identification of 2 novel genes (*PDE4DIP* and *ACOT4*) and the replication of 2 previously reported candidate genes (*ABCA1* and *ZFXH3*) as containing cod-

ing region variants associated with ischemic stroke. These new results suggest that 2 pathways, involving cell migration and growth (*PDE4DIP*) and long-chain fatty acid metabolism (*ACOT4*), could provide insights into the cause of ischemic stroke and as targets for pharmacologic interventions and therapies.

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REFERENCES

- Roger VL, Go AS, Lloyd-Jones DM, et al; American Heart Association Statistics Committee and Stroke Statistics Subcommittee. Executive summary: heart disease and stroke statistics—2012 update: a report from the American Heart Association. *Circulation*. 2012;125(1):188-197.
- Kim AS, Johnston SC. Temporal and geographic trends in the global stroke epidemic. *Stroke*. 2013; 44(6)(suppl 1):S123-S125.
- Vernooij MW, Ikram MA, Tanghe HL, et al. Incidental findings on brain MRI in the general population. *N Engl J Med*. 2007;357(18):1821-1828.
- Vermeer SE, Longstreth WT Jr, Koudstaal PJ. Silent brain infarcts: a systematic review. *Lancet Neurol*. 2007;6(7):611-619.
- Kiely DK, Wolf PA, Cupples LA, Beiser AS, Myers RH. Familial aggregation of stroke: the Framingham Study. *Stroke*. 1993;24(9):1366-1371.
- Schulz UG, Flossmann E, Rothwell PM. Heritability of ischemic stroke in relation to age, vascular risk factors, and subtypes of incident stroke in population-based studies. *Stroke*. 2004; 35(4):819-824.
- Meschia JF, Worrall BB, Rich SS. Genetic susceptibility to ischemic stroke. *Nat Rev Neurol*. 2011;7(7):369-378.
- Ariyaratnam R, Casas JP, Whittaker J, Smeeth L, Hingorani AD, Sharma P. Genetics of ischaemic stroke among persons of non-European descent: a meta-analysis of eight genes involving approximately 32,500 individuals. *PLoS Med*. 2007; 4(4):e131.
- Casas JP, Hingorani AD, Bautista LE, Sharma P. Meta-analysis of genetic studies in ischemic stroke: thirty-two genes involving approximately 18,000 cases and 58,000 controls. *Arch Neurol*. 2004;61 (11):1652-1661.

10. Holmes MV, Newcombe P, Hubacek JA, et al. Effect modification by population dietary folate on the association between MTHFR genotype, homocysteine, and stroke risk: a meta-analysis of genetic studies and randomised trials. *Lancet*. 2011;378(9791):584-594.
11. Meschia JF, Nalls M, Matarin M, et al; Siblings With Ischemic Stroke Study Investigators. Siblings With Ischemic Stroke Study: results of a genome-wide scan for stroke loci. *Stroke*. 2011;42(10):2726-2732.
12. Traylor M, Farrall M, Holliday EG, et al; Australian Stroke Genetics Collaborative, Wellcome Trust Case Control Consortium 2 (WTCCC2); International Stroke Genetics Consortium. Genetic risk factors for ischaemic stroke and its subtypes (the METASTROKE collaboration): a meta-analysis of genome-wide association studies. *Lancet Neurol*. 2012;11(11):951-962.
13. Holliday EG, Maguire JM, Evans TJ, et al; Australian Stroke Genetics Collaborative; International Stroke Genetics Consortium; Wellcome Trust Case Control Consortium 2. Common variants at 6p21.1 are associated with large artery atherosclerotic stroke. *Nat Genet*. 2012;44(10):1147-1151.
14. Bellenguez C, Bevan S, Gschwendtner A, et al; International Stroke Genetics Consortium (ISGC); Wellcome Trust Case Control Consortium 2 (WTCCC2). Genome-wide association study identifies a variant in HDAC9 associated with large vessel ischemic stroke. *Nat Genet*. 2012;44(3):328-333.
15. Manolio TA, Collins FS, Cox NJ, et al. Finding the missing heritability of complex diseases. *Nature*. 2009;461(7265):747-753.
16. Marth GT, Yu F, Indap AR, et al; 1000 Genomes Project. The functional spectrum of low-frequency coding variation. *Genome Biol*. 2011;12(9):R84.
17. Tennessen JA, Bigham AW, O'Connor TD, et al; Broad GO; Seattle GO; NHLBI Exome Sequencing Project. Evolution and functional impact of rare coding variation from deep sequencing of human exomes. *Science*. 2012;337(6090):64-69.
18. Cole JW, Stine OC, Liu X, et al. Rare variants in ischemic stroke: an exome pilot study. *PLoS One*. 2012;7(4):e35591. doi:10.1371/journal.pone.0035591.
19. Adams HP Jr, Bendixen BH, Kappelle LJ, et al. Classification of subtype of acute ischemic stroke. Definitions for use in a multicenter clinical trial. TOAST. Trial of Org 10172 in Acute Stroke Treatment. *Stroke*. 1993;24(1):35-41.
20. Wu MC, Lee S, Cai T, Li Y, Boehnke M, Lin X. Rare-variant association testing for sequencing data with the sequence kernel association test. *Am J Hum Genet*. 2011;89(1):82-93.
21. Li B, Leal SM. Methods for detecting associations with rare variants for common diseases: application to analysis of sequence data. *Am J Hum Genet*. 2008;83(3):311-321.
22. R Core Team. R: a language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria; 2012. <http://www.R-project.org/>. Accessed June 1, 2012.
23. Lange LA, Hu Y, Zhang H, et al; NHLBI Grand Opportunity Exome Sequencing Project. Whole-exome sequencing identifies rare and low-frequency coding variants associated with LDL cholesterol. *Am J Hum Genet*. 2014;94(2):233-245.
24. Gretarsdottir S, Thorleifsson G, Reynisdottir ST, et al. The gene encoding phosphodiesterase 4D confers risk of ischemic stroke. *Nat Genet*. 2003;35(2):131-138.
25. Meschia JF, Brown RD Jr, et al; SWISS Study Group; ISGS Study Group; MSGD Study Group. Phosphodiesterase 4D and 5-lipoxygenase activating protein in ischemic stroke. *Ann Neurol*. 2005;58(3):351-361.
26. Bevan S, Porteous L, Sitzer M, Markus HS. Phosphodiesterase 4D gene, ischemic stroke, and asymptomatic carotid atherosclerosis. *Stroke*. 2005;36(5):949-953.
27. Staton JM, Sayer MS, Hankey GJ, et al. Association between phosphodiesterase 4D gene and ischaemic stroke. *J Neurol Neurosurg Psychiatry*. 2006;77(9):1067-1069.
28. Worrall BB, Mychaleckyj JC. PDE4D and stroke: a real advance or a case of the emperor's new clothes? *Stroke*. 2006;37(8):1955-1957.
29. Yoon D, Park SK, Kang D, Park T, Park JW. Meta-analysis of homogeneous subgroups reveals association between PDE4D gene variants and ischemic stroke. *Neuroepidemiology*. 2011;36(4):213-222.
30. Bamshad MJ, Shendure JA, Valle D, et al; Centers for Mendelian Genomics. The Centers for Mendelian Genomics: a new large-scale initiative to identify the genes underlying rare mendelian conditions. *Am J Med Genet A*. 2012;158A(7):1523-1525.
31. Lee Y, Gamazon ER, Rebman E, et al. Variants affecting exon skipping contribute to complex traits. *PLoS Genet*. 2012;8(10):e1002998. doi:10.1371/journal.pgen.1002998.
32. Willett WC, Ascherio A. Trans fatty acids: are the effects only marginal? *Am J Public Health*. 1994;84(5):722-724.
33. Smit LA, Katan MB, Wanders AJ, Basu S, Brouwer IA. A high intake of trans fatty acids has little effect on markers of inflammation and oxidative stress in humans. *J Nutr*. 2011;141(9):1673-1678.
34. Mozaffarian D, Katan MB, Ascherio A, Stampfer MJ, Willett WC. Trans fatty acids and cardiovascular disease. *N Engl J Med*. 2006;354(15):1601-1613.
35. Lopez-Garcia E, Schulze MB, Meigs JB, et al. Consumption of trans fatty acids is related to plasma biomarkers of inflammation and endothelial dysfunction. *J Nutr*. 2005;135(3):562-566.
36. Kromhout D, Menotti A, Bloemberg B, et al. Dietary saturated and trans fatty acids and cholesterol and 25-year mortality from coronary heart disease: the Seven Countries Study. *Prev Med*. 1995;24(3):308-315.
37. Hunt MC, Rautanen A, Westin MA, Svensson LT, Alexson SE. Analysis of the mouse and human acyl-CoA thioesterase (ACOT) gene clusters shows that convergent, functional evolution results in a reduced number of human peroxisomal ACOTs. *FASEB J*. 2006;20(11):1855-1864.
38. Westin MA, Hunt MC, Alexson SE. The identification of a succinyl-CoA thioesterase suggests a novel pathway for succinate production in peroxisomes. *J Biol Chem*. 2005;280(46):38125-38132.
39. van Bilsen M, van Nieuwenhoven FA. PPARs as therapeutic targets in cardiovascular disease. *Expert Opin Ther Targets*. 2010;14(10):1029-1045.
40. Kilarski LL, Achterberg S, Devan WJ, et al; GARNET Collaborative Research Group, Wellcome Trust Case Control Consortium 2, Australian Stroke Genetic Collaborative, the METASTROKE Consortium, and the International Stroke Genetics Consortium. Meta-analysis in more than 17,900 cases of ischemic stroke reveals a novel association at 12q24.12. *Neurology*. 2014;83(8):678-685.